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The Role of Oxidative Stress in Metals Toxicity/ Mitochondrial Dysfunction as a Key Player

REVIEW

ARTICLE

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Abstract

Metals can cause oxidative stress by increasing the formation of reactive oxygen species (ROS), which make antioxidants incapable of defiance against growing amounts of free radicals. Metal toxicity is related to their oxidative state and reactivity with other compounds. However, several reports about metals have been published in the recent years. Mitochondria, as a site of cellular oxygen consumption and energy production, can be a target for metals toxicity. Dysfunction of Mitochondrial oxidative phosphorylation led to the production of some metals toxicities metals through alteration in the activities of I, II, III, IV and V complexes and disruption of mitochondrial membrane. Reductions of adenosine triphosphate (ATP) synthesis or induction of its hydrolysis can impair the cellular energy production. In the present review study, the researchers have criticized reviews and some evidence about the oxidative stress as a mechanism of toxicity of metals. The metals disrupt cellular and antioxidant defense, reactive oxygen species (ROS) generation, and promote oxidative damage. The oxidative injuries induced by metals can be restored by use of antioxidants such as chelators, vitamin E and C, herbal medicine, and through increasing the antioxidants level. However, to elucidate many aspect of mechanism toxicity of metals, further studies are yet to be carried out. **[GMJ. 2014;3(1):2-13]**

Keywords: Metals; Mitochondrial Dysfunction; Oxidative Stress; Reactive Oxygen Species

Introduction

Reactive oxygen species (ROS) are the most important group of radical species [1,2]. ROS and reactive nitrogen species (RNS) are known to play a dual role in biological organisms, since they can be either harmful or beneficial to living systems [3]. One further beneficial example of ROS at low concentrations is the induction mitogenic response. In contrast, at high concentrations, ROS can be important mediators of damage to cell structures, including lipids in cell mem-

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branes, proteins and nucleic acids [4]. Antioxidant actions of non-enzymatic antioxidants and antioxidant enzymes correct the adverse effects of ROS [5]. Oxidative stress can be defined most simply as the imbalance between the production of ROS, RNS capable of causing peroxidation of lipid layer of cells and the body's antioxidant defense [6].

There is some evidence showing metals such as iron, copper, cadmium, chromium, lead, mercury, nickel, vanadium and aluminum can produce ROS and RNS through lipid peroxidation, DNA damage, depletion of sulfhy-

Correspondence to: Akram Ranjbar, School of Pharmacy, Hamadan University of Medical Sciences, Hamadan, Iran Telephone Number: (+98) 811-8380031 Email Address : a.ranjbar@umsha.ac.ir dryls, and altered calcium homeostasis [7,8]. In metabolism of oxygen in aerobic organisms, the mitochondrial respiratory chain is the major source of intracellular ROS generation and at the same time, an important target for the damaging effects of ROS and RNS [9,10].

The interaction of diverse macromolecules with ROS/RNS may impair the function of these organelles and may directly influence cell viability and trigger cell death [10,11]. The decline of mitochondrial respiratory function may also be caused by damages to the effect of direct free radicals on proteins, lipids and other macromolecules, as well as the effects of the mutant oxidatively damaged in mtDNA [11,12]. The mitochondrial transcription could be sensitive to free radical attack, to lipid peroxidation products, or to both. It has been proposed that mitochondrial impairment being the result of oxidative-induced damage plays a critical role in the metals toxicity [13,14]. The overall objective of this paper is to provide a concise and current review of the effects of metals toxicity on mitochondrial function and oxidative stress.

Metals-induced oxidative stress

Free radicals are defined as atoms or molecules that contain one or more unpaired electrons; the toxicity of many xenobiotic, especially metals are associated with the production of free radicals, which are in turn toxicant and implicated in the pathophysiology of many diseases [6].

The possible role of oxidative damage in pathology of metals may contribute to their toxicity [15]. Increased rates at ROS generation have often been suggested to contribute to the toxicity of high levels of several other metals, including lead, cobalt, mercury, nickel, cadmium, molybdenum, vanadium, chromium and aluminum, as well as other elements such as selenium and arsenic [16]. However, the evidence for a primary role of oxidative stress in toxicity for the elements in question is not particularly convincing. For example, although increased lipid peroxidation has often been demonstrated in isolated cells exposed to metals, or in tissues from animals poisoned by metals, this peroxidation may be a consequence of tissue injury and GSH depletion cause by the metals rather than on early contributor to the metal toxicity [17,18]. Several studies have focused on metal-induced toxicity and carcinogenicity, emphasizing their role on the generation of ROS / RNS in biological systems [18].

Mitochondria: The major source cellular oxidative stress

As estimated [19], some 0.2–2% of the oxygen taken up by cells is converted by mitochondria to ROS, mainly through the production of superoxide anion. Mitochondria consume 85– 90% of a cell's oxygen to support oxidative phosphorylation, the major-energy production system in cells that works through oxidation of fuels through the synthesis adenosine triphosphate (ATP) [20].

Hence, the mitochondrial respiratory chain serves as a major source of ROS, derived from the disproportionation of superoxide anions [21]. Within mitochondria, it is the electron transport chain that is the main source of ROS [22]. The sites of ROS production along the chain have been subjected to many studies [23-25]. Recent findings show that two major sites of superoxide production are at complex I and complex III [26,27]. As described in previous studies [6,28,29], the term oxidative stress refers to both oxidative damage and oxidative stress impact on signaling, transcriptional control and other normal processes within cells; the term has also encompassed the effects of oxidants such as RNS.

In mammalian tissues, there are at least three distinct superoxide dismutase (SOD) isoenzymes, including one manganese form (Mn-SOD) present in the mitochondrial matrix and two copper and zinc forms (Cu,Zn-SOD), one of which is located only in the cytosol and the other one is in various extracellular fluids, respectively [30]. SOD plays a key role in catalyzing the dismutation of O2--to O2 and H2O2. Glutathione peroxidase (GPx) and catalase (CAT), remove hydrogen peroxide. In the presence of transition metals, H2O2 can be reduced to the extremely reactive OH [31,32]. Metabolizing water and corresponding alcohols (ROH) need to reduce H2O2 and a wide range of organic hydroperoxides (ROOH) by

some catalyzing reactions through GPx. Another abundant reactive radical is Nitric oxide (NO°). NO° acts as an important oxidative biological signaling molecule, having an important role on a large variety of diverse physiological processes. These include neurotransmission, blood pressure regulation, defense mechanisms, smooth muscle relaxation and immune regulation [33,34]. NO° is enzymatically generated by the actions of nitric oxide synthases (NOS) and has a half-life of only a few seconds in an aqueous environment. Under these conditions, Peroxynitrite anion ONOO° is an oxidizing free radical produced by NO° and the superoxide anion, being able to cause DNA fragmentation and lipid peroxidation [34,35].

Metal induced mitochondrial dysfunction and oxidative stress

Chromium

Chromium (Cr) is a chemical widely used in steel, alloy, cast, irons, chrome, paints, metal finishes and wood treatment. Cr is one of the important causes of allergic dermatitis and has toxic and carcinogenic effects on humans and animals [36-38]. Chromate plating and other hexavalent Cr (VI) exposure can occur in several industrial uses such as chromate pigments, chromate-based corrosion inhibitors, stainless steel machining and welding, etc [37,39]. The authors have reviewed recent in vitro and in vivo effects of oxygen scavengers, glutathione vitamin B2, vitamin E and vitamin C on chromate-induced injuries including DNA damage, lipid peroxidation, enzyme inhibition, cytotoxicity and mutagenesis. Also, Chromium overdoes occurs in the workplace primarily in the valence forms Cr (VI) and Cr (III) [40]. Inhalation of hexavalent chromium can result in several disorders such as pulmonary fibrosis, chronic bronchitis, lung cancer, occupational asthma and others [41-45]. Cr (VI) can also generate highly reactive oxidant such as peroxynitrite. In fact, Cr(VI) reduction results in several oxidants: (a) Cr(V) can directly oxidize cell components, (b) Cr(IV) catalyzes robust hydroxyl radical (HO°) generation in Fenton-like reactions with H2O2; and, (c) some enzymes simultaneously reduce Cr(VI)to Cr(V) and generate superoxide (O2°-) [46-48]. In previous studies, the results showed that Cr (VI) exposure significantly inhibits the activity of core mitochondrial functions (aconitase, complexes I and II) in both cultured cells and bronchial epithelium [49]. The inhibition of mitochondrial core protein results in inhibits of electron transfer chain and thereby impaired oxygen reduction. These phenomena lead to radicals' formation and oxidative stress [50]. The results of a study showed that total blood Cr level, SOD level, lipid peroxidation level and DNA damage were significantly higher and GSH level was significantly lower in exposed group as compared to the unexposed group [50]. Also, the studies showed that the toxicity of Cr (III) is mainly associated with cross linking mechanism which leads to multiform DNA damages, e.g., strand breakage, DNAprotein cross-links, DNA-DNA cross-links, Cr-DNA adducts and base modifications in cells [51-54]. Only chromium (VI) does not react with DNA in vitro, or in isolated nuclei. However, once inside the cell, in the presence of cellular reductants, it causes a wide variety of DNA lesions including Cr-DNA adducts, DNA-protein crosses links, DNA-DNA cross links and oxidative damage. Within the cell, glutathione rapidly forms a complex with Cr (VI), followed by a slow reduction of Cr (VI) to yield Cr (V). In addition, superoxide can further reduce Cr (VI) to Cr (V), which can further catalyze the demonstration of H2O2. Thus, it leads to the creation of DNA damaging hydroxyl radical [55,56]. Also, Cr is a ROS promoting agent, resulting in mitochondrial damage that leads to apoptosis and carcinogenicity. For example, in vivo, Cr (VI) exposure results in apoptosis, mitochondrial instability, release of cytochrome c and at least initiation cell disruption [57-59].

All in all, effective Cr chelating or elevated cellular antioxidation is the most useful way of treating Cr-induced oxidative injury, leading to the prevention of neurodegenerative disorders and chronic diseases [57,58].

Cadmium

Cadmium (Cd) is a highly toxic metal of occupational and environmental concern due to its widespread contamination of sites worldwide and long biological half-life (10 to 30 years) [59]. In Japan, Itai-Itai disease (sever Cd poisoning) was observed when Cd was discharged from a mine into a river used to supply drinking water. The organism is widely distributed in the environment and elevated exposure can be of both natural and anthropogenic origin [60,61]. Exposure occurs mainly via food, in particular plant-derived food and certain seafood, and from tobacco smoke [62]. Several studies showed that low-level environmental exposure to Cd has adverse health effect on kidney and bone; In addition, recent studies have reported higher risk of cancer and increased mortality [63]. In adults, only a few percentage of the ingested Cd is absorbed in the gastrointestinal tract. In contrast, Young adults have a higher absorption, apparently coupled with a different mechanism of uptake [64]. Cd-increased ROS lead to lipid peroxidation and DNA damage ROS has been implicated in chronic Cd nephrotoxicity [65-67], immunotoxicity [68] and carcinogenesis [69]. Some indirect mechanisms involve in radical production by Cd. Several mechanisms have emphasized the role of Cd in generation of free radicals. Disruption of the cellular antioxidant system by glutathione depletion is one of them [70,71]. Induction of inflammation in the liver is another important mechanism that proposed for Cd-induced oxidative stress [65]. Cd-induced inflammatory mediators such as IL-1 β , TNF- α , IL-6, and IL-8 are generated by the activation of the resident macrophages of the liver (Kupffer cells) [72]. It has been suggested that Cd produce ROS by binding to protein thiols in the mitochondrial membrane and affect mitochondrial permeability transition and inhibit respiratory chain reaction [73,74]. Cd inhibit mitochondrial complex III, resulting in accumulation of semiubiquinones at the Coenzyme Q sites, which lead to one electron to molecular oxygen to form superoxide anion [75]. Indeed, Cd effects on mitochondrial electron transfer are the major origin for Cd generated ROS, not only in mammalian cells, but also in plants [76]. In cells, some of transcription factors such as AP-1 and NF-kB are sensitive to oxidative stress. The activation of these

transcription factors by Cd has been shown in intact animals and cultured cells [77-79]. In addition, the activation of MAPKs by Cd is associated with ROS production in intact animals (80) in cultured cells [81,82], which in turn plays an important role in Cd-induced apoptosis to eliminate oxidative damaged cells [83]. It is hypothesized that during acute and chronic Cd exposure, adaptation mechanisms are induced to offset Cd-induced ROS, oxidative damage and mitochondrial dysfunction [70,71].

Adaptation to chronic Cd exposure reduces ROS production, but acquired Cd tolerance with aberrant gene expression plays important roles in acute, chronic Cd toxicity and apoptosis [77,84]. In addition, Cd modulates protein kinase, transcription factors, MAPK, mitochondria, caspases, and ROS pathways all seem to have a role in Cd-induced apoptosis and cancer [78,80]. Cooperatively, efficient chelation of the element and/or supplementing antioxidative materials is the preferred medical treatment for reducing various toxic and effects followed by Cd exposure [65,70].

Lead

Lead (Pb) is a common agent that causes environmental and industrial pollutant. Pb is one of the most commonly used metals in industry and its toxicity is of concern to public health due to the persistence of lead in the environment. Pb has been found to produce several toxic biochemicals [85].

Liver, kidney and brain are the major organ that affected by Pb [86]. Long term exposure to this bio-toxicant leads to its accumulation in these organs with maximum concentration in different tissues [87]. The neurotoxic effect of Pb, particularly in the developing brain is a matter of serious concern and behavioral abnormalities, learning impairment, decreased hearing and impaired cognitive functions in humans and experimental animals have been recorded with blood Pb levels as low as 10 g/ dl [88]. Several mechanisms have been proposed to explain the Pb induced toxicity, but no mechanisms have been yet defined explicitly [85]. Results from recent studies showed that oxidative stress is one of the important mechanisms of toxic effects of Pb [85]. Also,

Pb exposure led to various degrees of increased lipid peroxidation with tissue specific changes in liver [89,90], kidneys [89,91] and brain [89,92,93]. Treatment of Pb -exposed rats with tocopherol and ascorbic acid did not reduce tissue Pb burden, but lowered the lipid peroxidation levels, revealing their antioxidant potential in lead related oxidative stress. In addition Pb is shown to induce changes in the composition of red blood cell (RBC) membrane proteins and lipids, and inhibit hemoglobin synthesis [94,95]. Several antioxidant enzymes and molecules such as reduced glutathione (GSH) concentration, GPx, SOD and CAT activities, have been used to evaluate Pb -induced oxidative damage in animal and human studies [89,95,96]. Pb, because of its affinity to SH group, is known to inhibit ALAD (the second enzyme in the heme biosynthesis pathway and catalyzes condensation of two molecules of aminolevulinic acid (ALA) to a porphobilinogen.), resulting in accumulation of ALA. ALAD has been suggested as a sensitive index of the effect of Pb exposure on hematological system [97,98]. Hematological system is one of the important targets for Pb induced toxicity. The effects of Pb on this system result in decreased heme synthesis and anemia [99]. High concentrations of oxygen, autoxidizability of hemoglobin, vulnerable membrane components to lipid peroxidation and limited capacity to repair their damaged components, are factors that make RBCs sensitive to oxidative damage [100]. Previous studies of correlation between clinical indicators of Pb poisoning and oxidative stress parameters in controls and Pb-exposed workers showed that there was a disruption of prooxidant/antioxidant balance in Pb-exposed workers [101,102]. However, several studies suggested ALA as a possible source [103]. Inhibition of ALAD by lead results in accumulation of ALA during heme biosynthesis. In next step, accumulated ALA has been shown to undergo metal-catalyzed auto-oxidation giving rise to the formation of superoxide (O2•-), H2O2 and ALA [103]. All in all, there may be two independent sources of Pb-induced oxidative damage; the first is the pro-oxidative effect of δ -ALA, and the second is connected with the direct effect of Pb on membrane lip-

ids and mitochondrial dysfunction [104-106]. Pb depolarizes cell mitochondria due to the opening of permeability transition pore, resulting in cytochrome c release, caspase activation, and apoptosis. In Pb induced apoptosis, the opening of mitochondrial permeability transition pore is due to oxidative stress [107,108].

There are many studies suggesting possible clinical applications of exogenous antioxidants in the treatment of toxicity induced by Pb exposure. For example, treatment with ascorbic acid or a-tocopherol and N-acetylcystein was found to reduce the level of ROS-initiated damage and their combined administration restored normal mitochondrial function in Pb -supplemented rats [89,109,110].

Aluminum

Aluminum (Al) is the third most abundant element and distributed widely in the biosphere. Al constitutes approximately 8% of the earth crust exceeded only by oxygen #7% and silicon 28% [8]. Several mechanisms have been proposed to explain the toxicity of Al, none supported by convincing data from in vivo experiments [111]. Al3+ ions cannot stimulate lipid peroxidation or other free radical reactions which is not surprising because of their fixed valence [112]. However, if peroxidation in liposomes erythrocytes, synaptosome somylein, or microsomes is stimulated by adding fe2+ ions, the simultaneous addition of Al3+ increases the per oxidation rate. It may be that Al3+ ions bind to membranes and cause a subtle rearrangement of membrane lipids that aids the propagation of lipid per oxidation, this action of Al3+ might contribute to its neurotoxin properties, since the brain is sensitive to oxidative damage [113,114]. Injection of large dose of aluminum salts into animals has been claimed to increase brain lipid peroxidation levels. And injection of aluminum-containing vaccines into mice caused a transient rise in brain aluminum levels [114,115].

Also, Aluminum accumulation is thought to be related to renal impairment, anemia and other clinical complications in hemodialysis patients and they showed that patients undergoing hemodialysis present platelet dysfunction and lipid peroxidation [116,117].

However, increased ROS were reported during Al exposure, which was attributed to electron leakage, enhanced mitochondrial activity and increased electron chain activity. Mitochondria contribute too much of core human metabolism, including oxidative phosphorylation, the tricarboxylic acid (TCA) cycle, fatty acid oxidation, iron sulfur center and heme biosynthesis, and amino acid metabolism [118]. These are in addition to the well-established role of the mitochondria in energy metabolism and regulation of cell death [119]. Also, previous studies revealed that Al induced imbalance in this steady state allows the induction and effects of mitochondrial dysfunction [120,121]. Since Al induces oxidative damage resulting in an increase of ROS production, it is possible that Al-induced ROS are involved in mitochondrial instability, and release of cytochrome c [122,123]. Although the administration of antioxidant materials is widely used in Al intoxication. vitamin C, E and efficient chelating of the element supplementing is useful, that Al induced, neurode generative disorders such as Alzheimer diseases [124-127].

Conclusion

One cannot avoid the generation of ROS, because it is a result from aerobic life. ROS is produced in mitochondrial function [128]. ROS are known not only to attack DNA, but additional cellular components \$uch as proteins and lipids, leaving after reactive species that can, in turn, bind to DNA bases [129,130]. This implicates \$uch damage in the etiology of many diseases such as cancer [131,132]. Toxic metals (lead, ICd, Cr, Al, mercury and arsenic) are widely found in our environment [17].

Humans are exposed to these metals from numerous sources, including contaminated air, water, soil and food [16]. There are some new studies, showing that transition metals act as catalysts in the oxidative reactions of biological macromolecules. So, metals act

their toxicities roles through mitochondrial dysfunction and oxidative tissue damage [16,18,133,134]. Although, ROS has been implicated in activation of an extrinsic cascade, the correlation of ROS, caspase activation and p38 in metals-induced apoptosis, requires further investigation. A potential role for ROS, the mitochondria, and activation of several signaling pathways (MAPK and p53) have been established for several metals [135,136]. To clarify how ROS induce cellular response and signal transduction is quite important for understanding of the mechanisms of metal-induced carcinogenesis. Certainly, many researchers have implicated the involvement of ROS signaling in metal-induced carcinogenesis and cell death over the last decade. However, they did not provide a direct evidence of the correlation between ROS and metal-induced apoptosis and carcinogenesis [137-139].

Data suggest that antioxidants may play a fimportant role in abating some hazards of metals [8,140]. Currently, treatments against metals toxicity include the use of chelating agents, metallothionein, and antioxidant therapy with melatonin, vitamin E, vitamin C, N-acetylcystein and herbal medicine [7,141-144]. The effectiveness of an antioxidant based treatment approach is dependent on understanding the mechanisms by which metals cause mitochondrial dysfunction and other health conditions. Although metal-induced oxidative stress does not explain all of the cell disruptions' caused by metals, accumulating evidence emphasizes the protective effect of antioxidants in the setting of metal-induced toxicity. The effectiveness of an antioxidant-based treatment approach is dependent on understanding the mechanisms by which metals cause cancer and other health conditions. In this regard, future studies should focus on defining cellular and molecular mechanisms of metal-induced oxidative injuries, developing efficient biomarkers, and identifying individuals with increased susceptibility to metal exposure.

References

- Halliwell B. Biochemistry of oxidative stress. Biochemical Society Transactions. Biochem Soc Trans. 2007;35(Pt 5):1147-50.
- Ranjbar A, Khorami S, Safarabadi M, Shahmoradi A, Malekirad AA, Vakilian K, et al. Antioxidant activity of Iranian Echium amoenum Fisch & CA Mey flower decoction in humans: a cross-sectional before/after clinical trial. Evid Based Complement Alternat Med. 2006;3(4):469-73.
- Guo T, Cui L, Shen J, Wang R, Zhu W, Xu Y, et al. A dual-emission and large Stokes shift fluorescence probe for real-time discrimination of ROS/RNS and imaging in live cells. Chem Commun (Camb). 2013;49(18):1862-4.
- Dröge W. Free radicals in the physiological control of cell function. Physiol Rev. 2002;82(1):47-95.
- Halliwell B, Gutteridge J, Cross C. Free radicals, antioxidants, and human disease: where are we now? J Lab Clin Med.. 1992;119(6):598-620.
- Mittler R. Oxidative stress, antioxidants and stress tolerance. Trends Plant Sci. 2002;7(9):405-10.
- Valko M, Rhodes C, Moncol J, Izakovic M, Mazur M. Free radicals, metals and antioxidants in oxidative stress-induced cancer. Chem Biol Interact. 2006;160(1):1-40.
- Ranjbar A, Khani-Jazani R, Sedighi A, Jalali-Mashayekhi F, Ghazi-Khansari M, Abdollahi M. Alteration of body total antioxidant capacity and thiol molecules in human chronic exposure to aluminum. Toxicological & Environmental Chemistry. 2008;90(4):707-13.
- Cadenas E, Davies KJ. Mitochondrial free radical generation, oxidative stress, and aging. Free Radic Biol Med. 2000;29(3-4):222-30.
- Ott M, Gogvadze V, Orrenius S, Zhivotovsky B. Mitochondria, oxidative stress and cell death. Apoptosis. 2007;12(5):913-22.
- Orrenius S, Gogvadze V, Zhivotovsky B. Mitochondrial oxidative stress: implications for cell death. Annu Rev Pharmacol Toxicol. 2007;47:143-83.
- 12. Golden T-R, Melov S. Mitochondrial DNA mutations, oxidative stress, and aging. Mech

Ageing Dev. 2001;122(14):1577-89.

- Lin MT, Beal MF. Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. Nature. 2006;443(7113):787-95.
- 14. Krumschnabel G, Manzl C, Berger C, Hofer B. Oxidative stress, mitochondrial permeability transition, and cell death in Cu-exposed trout hepatocytes. Toxicol Appl Pharmacol. 2005;209(1):62-73.
- Huang X, Moir RD, Tanzi RE, Bush AI, Rogers JT. Redox-Active Metals, Oxidative Stress, and Alzheimer's Disease Pathology. Ann N Y Acad Sci. 2004;1012(1):153-63.
- Valko M, Morris H, Cronin M. Metals, toxicity and oxidative stress. Curr Med Chem. 2005;12(10):1161-208.
- Ercal N, Gurer-Orhan H, Aykin-Burns N. Toxic metals and oxidative stress part I: mechanisms involved in metal-induced oxidative damage. Curr Top Med Chem. 2001;1(6):529-39.
- Jomova K, Valko M. Advances in metalinduced oxidative stress and human disease. Toxicology. 2011;283(2):65-87.
- Hansford RG, Hogue BA, Mildaziene V. Dependence of H2O2 formation by rat heart mitochondria on substrate availability and donor age. J Bioenerg Biomembr. 1997;29(1):89-95.
- Requejo R, Chouchani ET, Hurd TR, Menger KE, Hampton MB, Murphy MP. Measuring mitochondrial protein thiol redox state. Methods in enzymology. Methods Enzymol. 2010;474:123-47.
- Brand MD. The sites and topology of mitochondrial superoxide production. Experimental gerontology. Exp Gerontol. 2010;45(7):466-72.
- 22. Ranjbar A, Ghahremani MH, Sharifzadeh M, Golestani A, Ghazi-Khansari M, Baeeri M, et al. Protection by pentoxifylline of malathion-induced toxic stress and mitochondrial damage in rat brain. Hum Exp Toxicol. 2010;29(10):851-64.
- 23. Brand M. Uncoupling to survive? The role of mitochondrial inefficiency in ageing. Exp Gerontol. 2000;35(6):811-20.
- Starkov AA, Fiskum G, Chinopoulos C, Lorenzo BJ, Browne SE, Patel MS, et al. Mitochondrial α-ketoglutarate dehydrogenase complex generates

reactive oxygen species. J Neurosci. 2004;24(36):7779-88.

- 25. Jang YC, Lustgarten MS, Liu Y, Muller FL, Bhattacharya A, Liang H, et al. Increased superoxide in vivo accelerates age-associated muscle atrophy through mitochondrial dysfunction and neuromuscular junction degeneration. The FASEB J. 2010;24(5):1376-90.
- 26. Weisiger RA, Fridovich I. Mitochondrial superoxide dismutase site of synthesis and intramitochondrial localization. J Biol Chem. 1973;248(13):4793-6.
- Liang L, Ho Y, Patel M. Mitochondrial superoxide production in kainate-induced hippocampal damage. Neuroscience. 2000;101(3):563-70.
- Sies H, Cadenas E. Oxidative stress: damage to intact cells and organs. Philos Trans R Soc Lond B Biol Sci. 1985;311(1152):617-31.
- 29. Abdollahi M, Ranjbar A, Shadnia S, Nikfar S, Rezaie A. Pesticides and oxidative stress: a review. Med Sci Monit. 2004;10(6):RA141-7.
- Young I, Woodside J. Antioxidants in health and disease. J Clin Pathol. 2001;54(3):176-86.
- Yon J-M, Baek I-J, Lee S-R, Jin Y, Kim M-R, Nahm S-S, et al. The spatio-temporal expression pattern of cytoplasmic Cu/ Zn superoxide dismutase (SOD1) mRNA during mouse embryogenesis. J Mol Histol. 2008;39(1):95-103.
- 32. Fantel AG, Mackler B, Stamps LD, Tran TT, Person RE. Reactive oxygen species and DNA oxidation in fetal rat tissues. Free Radic Biol Med. 1998;25(1):95-103.
- Flohé L. Glutathione peroxidase. Oxygen Radicals in Biology and Medicine. Springer US; 1988. Volume,49. p. 663-8.
- Flohé L. Glutathione Peroxidases. Selenoproteins and Mimics. Springer Berlin Heidelberg; 2012. p. 1-25.
- Webb AJ, Patel N, Loukogeorgakis S, Okorie M, Aboud Z, Misra S, et al. Nitric Oxide, Oxidative Stress. Hypertension. 2008;51(3):784-90.
- Stohs SJ, Bagchi D, Hassoun E, Bagchi M. Oxidative mechanisms in the toxicity of chromium and cadmium ions. J Environ Pathol Toxicol Oncol. 2001;20(2):77-88.
- Barnhart J. Occurrences, uses, and properties of chromium. Regul Toxicol Pharmacol. 1997;26(1):S3-7.

- 38. Vincent J. The nutritional biochemistry of chromium (III). 1st ed. Elsevier; 2011.
- Bagchi D, Bagchi M, Stohs SJ. Chromium (VI)-induced oxidative stress, apoptotic cell death and modulation of p53 tumor suppressor gene. Molecular Mechanisms of Metal Toxicity and Carcinogenesis: Springer; 2001. p. 149-58.
- Mertz W. Chromium occurrence and function in biological systems. Physiol Rev. 1969;49(2):163-239.
- 41. Anderson RA. Chromium metabolism and its role in disease processes in man. Clin Physiol Biochem. 1985;4(1):31-41.
- 42. Wedeen RP, Qian L. Chromium-induced kidney disease. Environ Health Perspect. 1991;92:71-4.
- Hummel M, Standl E, Schnell O. Chromium in metabolic and cardiovascular disease. Horm Metab Res. 2007;39(10):743-51.
- 44. Wang ZQ, Cefalu WT. Current concepts about chromium supplementation in type 2 diabetes and insulin resistance. Curr Diab Rep.. 2010;10(2):145-51.
- 45. Gambelunghe A, Piccinini R, Ambrogi M, Villarini M, Moretti M, Marchetti C, et al. Primary DNA damage in chrome-plating workers. Toxicology. 2003;188(2):187-95.
- Newbold R, Amos J, Connell J. The cytotoxic, mutagenic and clastogenic effects of chromium-containing compounds on mammalian cells in culture. Mutat Res. 1979;67(1):55-63.
- Levis A, Bianchi V, Tamino G, Pegoraro B. Cytotoxic effects of hexavalent and trivalent chromium on mammalian cells in vitro. Br J Cancer. 1978;37(3):386-96.
- Vasylkiv OY, Kubrak OI, Storey KB, Lushchak VI. Cytotoxicity of chromium ions may be connected with induction of oxidative stress. Chemosphere. 2010;80(9):1044-9.
- Rossi SC, Gorman N, Wetterhahn KE. Mitochondrial reduction of the carcinogen chromate: formation of chromium (V). Chem Res Toxicol. 1988;1(2):101-7.
- 50. Khan FH, Ambreen K, Fatima G, Kumar S. Assessment of health risks with reference to oxidative stress and DNA damage in chromium exposed population. Sci Total Environ. 2012;430:68-74.
- Bagchi D, Stohs SJ, Downs BW, Bagchi M, Preuss HG. Cytotoxicity and oxidative mechanisms of different forms of chromium. Toxicology. 2002;180(1):5-22.

- Peterson-Roth E, Reynolds M, Quievryn G, Zhitkovich A. Mismatch repair proteins are activators of toxic responses to chromium-DNA damage. Mol Cell Biol. 2005;25(9):3596-607.
- Sugden KD, Stearns DM. The role of chromium (V) in the mechanism of chromate-induced oxidative DNA damage and cancer. J Environ Pathol Toxicol Oncol. 2000;19(3):215-30.
- 54. Arakawa H, Weng M-w, Chen W-c, Tang M-s. Chromium (VI) induces both bulky DNA adducts and oxidative DNA damage at adenines and guanines in the p53 gene of human lung cells. Carcinogenesis. 2012;33(10):1993-2000.
- 55. Chiu A, Shi X, Lee W, Hill R, Wakeman T, Katz A, et al. Review of chromium (VI) apoptosis, cell-cycle-arrest, and carcinogenesis. J Environ Sci Health C Environ Carcinog Ecotoxicol Rev. 2010;28(3):188-230.
- 56. Aiyar J, Borges KM, Floyd RA, Wetterhahn KE. Role of chromium (V), glutathione thiyl radical and hydroxyl radical intermediates in Chromium (VI)-induced DNA Damage. Environ Health Perspect. 1991;92:53-62.
- 57. Sahin K, Sahin N, Kucuk O. Effects of chromium, and ascorbic acid supplementation on growth, carcass traits, serum metabolites, and antioxidant status of broiler chickens reared at a high ambient temperature (32 C). Nutr Res. 2003;23(2):225-38.
- Guha G, Rajkumar V, Kumar RA, Mathew L. Antioxidant activity of Lawsonia inermis extracts inhibits chromium (VI)-induced cellular and DNA toxicity. Evid Based Complement Alternat Med. 2011;2011:576456.
- Friberg L, Piscator M, Nordberg GF, Kjellström T. Cadmium in the environment. In: Sankar Das M. Trace analysis and technological development. International Union of Pure and Applied Chemistry; 1981.
- Kasuya M. Recent epidemiological studies on itai-itai disease as a chronic cadmiumpoisoning in Japan. Water Science & Technology. 2000;42(7-8):147-54.
- 61. Murata I, Hirono T, Saeki Y, Nakagawa S. Cadmium enteropathy, renal osteomalacia (" Itai Itai" disease in Japan). Bull Soc Int Chir. 1970;29(1):34-42.
- 62. Oskarsson A, Widell A, Olsson M, Grawé KP. Cadmium in food chain and health

effects in sensitive population groups. Biometals. 2004;17(5):531-4.

- 63. Satarug S, Garrett SH, Sens MA, Sens DA. Cadmium, environmental exposure, and health outcomes. Cien Saude Colet. 2011;16(5):2587-602.
- 64. Shim J, Son Y, Park JM, Kim MK. Effect of Chlorella intake on Cadmium metabolism in rats. Nutr Res Pract. 2009;3(1):15-22.
- 65. Shaikh ZA, Vu TT, Zaman K. Oxidative stress as a mechanism of chronic cadmiuminduced hepatotoxicity and renal toxicity and protection by antioxidants. Toxicol Appl Pharmacol. 1999;154(3):256-63.
- Prozialeck WC, Edwards JR, Vaidya VS, Bonventre JV. Preclinical evaluation of novel urinary biomarkers of cadmium nephrotoxicity. Toxicol Appl Pharmacol. 2009;238(3):301-5.
- A Shaikh Z, Tang W. Protection against chronic cadmium toxicity by glycine. Toxicology. 1999;132(2):139-46.
- Zhang X, Li D, Dong S, Zhang J. Study on the relationship between cadmium immunotoxicity and corticotropin-releasing factor. Wei sheng yan jiu. 2000;29(4):193-6.
- 69. Waalkes MP. Cadmium carcinogenesis. Mutat Res. 2003;533(1):107-20.
- Cuypers A, Plusquin M, Remans T, Jozefczak M, Keunen E, Gielen H, et al. Cadmium stress: an oxidative challenge. Biometals. 2010;23(5):927-40.
- Liu J, Qu W, Kadiiska MB. Role of oxidative stress in cadmium toxicity and carcinogenesis. Toxicol Appl Pharmacol. 2009;238(3):209-14.
- Rikans LE, Yamano T. Mechanisms of cadmium-mediated acute hepatotoxicity. J Biochem Mol Toxicol. 2000;14(2):110-7.
- Al-Nasser IA. Cadmium hepatotoxicity and alterations of the mitochondrial function. J Toxicol Clin Toxicol. 2000;38(4):407-13.
- 74. Li M, Xia T, Jiang CS, Li LJ, Fu JL, Zhou ZC. Cadmium directly induced the opening of membrane permeability pore of mitochondria which possibly involved in cadmium-triggered apoptosis. Toxicology. 2003;194(1):19-33.
- 75. Wang Y, Fang J, Leonard SS, Krishna Rao KM. Cadmium inhibits the electron transfer chain and induces reactive oxygen species. Free Radic Biol Med. 2004;36(11):1434-43.
- 76. Heyno E, Klose C, Krieger-Liszkay A. Origin of cadmium-induced reactive oxygen species production: mitochondrial electron

transfer versus plasma membrane NADPH oxidase. New Phytologist. 2008;179(3):687-99.

- Azzouzi BE, Tsangaris GT, Pellegrini O, Manuel Y, Benveniste J, Thomas Y. Cadmium induced apoptosis in a human T cell line. Toxicology. 1994;88(1):127-39.
- Kondoh M, Araragi S, Sato K, Higashimoto M, Takiguchi M, Sato M. Cadmium induces apoptosis partly via caspase-9 activation in HL-60 cells. Toxicology. 2002;170(1):111-7.
- Wang C, Ma W, Su Y. NF-κB Pathway Contributes to Cadmium-Induced Apoptosis of Porcine Granulosa Cells. Biol Trace Elem Res. 2013;153(1-3):403-10.
- Zhang C, Mao W, Kong X, Yue L, Gao Y, Yin Z. Inhibition of cadmium-induced apoptosis by Glutathione S-transferase P1 via mitogen-activated protein kinases (MAPKs) and mitochondria. Environ Toxicol Pharmacol. 2010;30(2):202-8.
- Fujiki K, Inamura H, Matsuoka M. Phosphorylation of FOXO3a by PI3K/Akt pathway in HK-2 renal proximal tubular epithelial cells exposed to cadmium. Arch Toxicol. 2013;87(12):2119-27.
- Nguyen KC, Willmore WG, Tayabali AF. Cadmium telluride quantum dots cause oxidative stress leading to extrinsic and intrinsic apoptosis in hepatocellular carcinoma HepG2 cells. Toxicology. 2013;306:114-23.
- Yang Z, Yang S, Qian SY, Hong J-S, Kadiiska MB, Tennant RW, et al. Cadmiuminduced toxicity in rat primary mid-brain neuroglia cultures: role of oxidative stress from microglia. Toxicol Sci. 2007;98(2):488-94.
- Brama M, Politi L, Santini P, Migliaccio S, Scandurra R. Cadmium-induced apoptosis and necrosis in human osteoblasts: role of caspases and mitogen-activated protein kinases pathways. J Endocrinol Invest. 2012;35(2):198-208.
- 85. Patrick L. Lead toxicity, a review of the literature. Part I: exposure, evaluation, and treatment. Altern Med Rev. 2006;11(1):2-22.
- 86. Gidlow D. Lead toxicity. Occup Med (Lond). 2004;54(2):76-81.
- Humphreys D. Effects of exposure to excessive quantities of lead on animals. Br Vet J. 1991;147(1):18-30.
- Sanders T, Liu Y, Buchner V, Tchounwou PB. Neurotoxic effects and biomarkers of lead exposure: a review. Rev Environ

Health. 2009;24(1):15-46.

- Patra R, Swarup D, Dwivedi S. Antioxidant effects of α tocopherol, ascorbic acid and L-methionine on lead induced oxidative stress to the liver, kidney and brain in rats. Toxicology. 2001;162(2):81-8.
- Sandhir R, Gill K. Effect of lead on lipid peroxidation in liver of rats. Biol Trace Elem Res. 1995;48(1):91-7.
- Wang L, Li J, Li J, Liu Z. Effects of lead and/or cadmium on the oxidative damage of rat kidney cortex mitochondria. Biological trace element research. Biol Trace Elem Res. 2010 Oct;137(1):69-78.
- 92. Prasanthi R, Devi CB, Basha DC, Reddy NS, Reddy GR. Calcium and zinc supplementation protects lead (Pb)-induced perturbations in antioxidant enzymes and lipid peroxidation in developing mouse brain. Int J Dev Neurosci. 2010;28(2):161-7.
- 93. Villeda-Hernandez J, Barroso-Moguel R, Mendez-Armenta M, Nava-Ruiz C, Huerta-Romero R, Rios C. Enhanced brain regional lipid peroxidation in developing rats exposed to low level lead acetate. Brain Res Bull. 2001;55(2):247-51.
- 94. Buettner GR. The pecking order of free radicals and antioxidants: lipid peroxidation, α-tocopherol, and ascorbate. Arch Biochem Biophys.1993;300(2):535-43.
- 95. Daniel EE. Ameliorative Effect of Vitamin C on Serum Liver Enzymes in Lead-Induced Toxicity in Wistar Rats. J Sci. 2013;3(1):188-912.
- 96. Bokara KK, Brown E, McCormick R, Yallapragada PR, Rajanna S, Bettaiya R. Lead-induced increase in antioxidant enzymes and lipid peroxidation products in developing rat brain. Biometals. 2008;21(1):9-16.
- 97. Sugawara E, Nakamura K, Miyake T, Fukumura A, Seki Y. Lipid peroxidation and concentration of glutathione in erythrocytes from workers exposed to lead. Br J Ind Med. 1991;48(4):239-42.
- 98. Goyer RA, Clarkson TW. Toxic effects of metals. Casarett & Doull's Toxicology The Basic Science of Poisons. 5th ed. Klaassen, CD [Ed], McGraw-Hill: Health Professions Division; 2011.
- 99. Ullah N, Khan MF, Mukhtiar M, Khan H, Rehman AU. Metabolic modulation of glutathione in whole blood components against lead-induced toxicity. African J Biotechnol. 2011;10(77):17853-8.

- 100. Campana O, Sarasquete C, Blasco J. Effect of lead on ALA-D activity, metallothionein levels, and lipid peroxidation in blood, kidney, and liver of the toadfish< Halobatrachus didactylus. Ecotoxicol Environ Saf. 2003;55(1):116-25.
- 101. Jiun YS, Hsien LT. Lipid peroxidation in workers exposed to lead. Arch Environ Health. 1994;49(4):256-9.
- 102. Gurer-Orhan H, Sabir HU, Özgüneş H. Correlation between clinical indicators of lead poisoning and oxidative stress parameters in controls and lead-exposed workers. Toxicology. 2004;195(2):147-54.
- 103. Rocha JB, Pereira ME, Emanuelli T, Christofari RS, Souz DO. Effect of treatment with mercury chloride and lead acetate during the second stage of rapid postnatal brain growth on δ-aminolevulinic acid dehydratase (ALA-D) activity in brain, liver, kidney and blood of suckling rats. Toxicology. 1995;100(1):27-37.
- 104. Rodriguez BL, Curb JD, Davis J, Shintani T, Perez MH, Apau-Ludlum N, et al. Use of the Dietary Supplement 5-Aminiolevulinic Acid (5-ALA) and Its Relationship with Glucose Levels and Hemoglobin A1C among Individuals with Prediabetes. Clin Transl Sci. 2012;5(4):314-20.
- 105. Ahamed M, Siddiqui M. Low level lead exposure and oxidative stress: current opinions. Clin Chim Acta. 2007;383(1):57-64.
- 106. Young KW, Piñon LG, Bampton ET, Nicotera P. Different pathways lead to mitochondrial fragmentation during apoptotic and excitotoxic cell death in primary neurons. J Biochem Mol Toxicol. 2010;24(5):335-41.
- 107. He L, Poblenz AT, Medrano CJ, Fox DA. Lead and calcium produce rod photoreceptor cell apoptosis by opening the mitochondrial permeability transition pore. J Biol Chem. 2000;275(16):12175-84.
- 108. Liu Z, Li D, Zhao W, Zheng X, Wang J, Wang E. A potent lead induces apoptosis in pancreatic cancer cells. PloS one. 2012;7(6):e37841.
- 109. Hsu P-C, Guo YL. Antioxidant nutrients and lead toxicity. Toxicology. 2002;180(1):33-44.
- 110. Gargouri M, Magné C, Dauvergne X, Ksouri R, El Feki A, Metges M-AG, et al. Cytoprotective and antioxidant effects of the edible halophyte Sarcocornia perennis

L.(swampfire) against lead-induced toxicity in renal cells. Ecotoxicol Environ Saf. 2013;95:44-51.

- 111.Berlyne G, Ben Ari J, Knopf E, Yagil R, Weinberger G, Danovitch G. Aluminium toxicity in rats. Lancet. 1972;299(7750):564-8.
- 112. Boscolo PR, Menossi M, Jorge RA. Aluminum-induced oxidative stress in maize. Phytochemistry. 2003;62(2):181-9.
- 113. Xie CX, Mattson MP, Lovell MA, Yokel RA. Intraneuronal aluminum potentiates iron-induced oxidative stress in cultured rat hippocampal neurons. Brain Res. 1996;743(1):271-7.
- 114. Sood PK, Nahar U, Nehru B. Curcumin attenuates aluminum-induced oxidative stress and mitochondrial dysfunction in rat brain. Neurotox Res. 2011;20(4):351-61.
- 115. Flora SJ, Mehta A, Satsangi K, Kannan GM, Gupta M. Aluminum-induced oxidative stress in rat brain: Response to combined administration of citric acid and HEDTA. Comp Biochem Physiol C Toxicol Pharmacol. 2003;134(3):319-28.
- 116. Mahieu ST, Gionotti M, Millen N, Elías MM. Effect of chronic accumulation of aluminum on renal function, cortical renal oxidative stress and cortical renal organic anion transport in rats. Arch Toxicol. 2003;77(11):605-12.
- 117. Wang H, Liang W, Huang J. Putrescine Mediates Aluminum Tolerance in Red Kidney Bean by Modulating Aluminum-Induced Oxidative Stress. Crop Science. 2013;53(5):2120-8.
- 118. Federico A, Cardaioli E, Da Pozzo P, Formichi P, Gallus GN, Radi E. Mitochondria, oxidative stress and neurodegeneration. J Neurol Sci. 2012;322(1):254-62.
- 119. Niemi NM, MacKeigan JP. Mitochondrial Phosphorylation in Apoptosis: Flipping the Death Switch. Antioxid Redox Signal. 2013;19(6):572-82.
- 120. Li Z, Xing D. Mechanistic study of mitochondria-dependent programmed cell death induced by aluminium phytotoxicity using fluorescence techniques. J Exp Bot. 2011;62(1):331-43..
- 121.Kumar V, Bal A, Gill KD. Impairment of mitochondrial energy metabolism in different regions of rat brain following chronic exposure to aluminium. Brain Res. 2008;1232:94-103.

- 122. Bharathi VP, Govindaraju M, Palanisamy A, Sambamurti K, Rao K. Molecular toxicity of aluminium in relation to neurodegeneration. Indian J Med Res. 2008;128(4):545-56.
- 123. Schildknecht PH, Vidal BC. Aluminium triggers necrosis and apoptosis in V79 cells. Toxicol Enviro Chem. 2004;86(1):63-72.
- 124. Gupta VB, Anitha S, Hegde M, Zecca L, Garruto R, Ravid R, et al. Aluminium in Alzheimer's disease: are we still at a crossroad? CMLS. 2005;62(2):143-58.
- 125. El-Demerdash FM. Antioxidant effect of vitamin E and selenium on lipid peroxidation, enzyme activities and biochemical parameters in rats exposed to aluminium. J Trace Elem Med Biol. 2004;18(1):113-21.
- 126. Silva S, Pinto G, Correia B, Pinto-Carnide O, Santos C. Rye oxidative stress under long term Al exposure. J Plant Physiol. 2013;170(10):879-89.
- 127. Rottkamp CA, Nunomura A, Raina AK, Sayre LM, Perry G, Smith MA. Oxidative stress, antioxidants, and Alzheimer disease. Alzheimer Dis Assoc Disord. 2000;14(1):S62-6.
- 128. Zorov DB, Juhaszova M, Sollott SJ. Mitochondrial ROS-induced ROS release: an update and review. Biochim Biophys Acta. 2006;1757(5):509-17.
- 129. Blokhina O, Virolainen E, Fagerstedt KV. Antioxidants, oxidative damage and oxygen deprivation stress: a review. Ann Bot. 2003;91(2):179-94.
- 130. Klaunig JE, Kamendulis LM, Hocevar BA. Oxidative stress and oxidative damage in carcinogenesis. Toxicol pathol. 2010;38(1):96-109.
- 131. Halliwell B, Gutteridge J. Free radicals in biology and medicine. Clarendon Press, Pergamon; 1985.
- 132. Costa M. Dioxygenases as Targets of Metals, Hypoxia and Oxidative Stress during Carcinogenesis. J Mol Genet Med. 2013.
- 133. Bahavar M, Tarbali N, Einolahi N, Dashti N. Evaluation of trace metal (cd, cr, cu)– induced oxidative stress in presence of H2O2 on purified DNA strands break from

nonpathogenic Escherichia coli. KAUMS Journal (FEYZ). 2013;16(7):633-4.

- 134. Belyaeva EA, Sokolova TV, Emelyanova LV, Zakharova IO. Mitochondrial electron transport chain in heavy metal-induced neurotoxicity: effects of cadmium, mercury, and copper. ScientificWorldJournal. 2012;2012:136063.
- 135. Pulido MD, Parrish AR. Metal-induced apoptosis: mechanisms. Mutat Res. 2003;533(1):227-41.
- 136. Granchi D, Cenni E, Ciapetti G, Savarino L, Stea S, Gamberini S, et al. Cell death induced by metal ions: necrosis or apoptosis? J Mater Sci Mater Med. 1998;9(1):31-7.
- 137. Shi H, Hudson LG, Liu KJ. Oxidative stress and apoptosis in metal ion-induced carcinogenesis. Free Radic Biol Med. 2004;37(5):582-93.
- 138. Kasprzak KS. Possible role of oxidative damage in metal-induced carcinogenesis. Cancer invest. 1995;13(4):411-30.
- 139. Martinez-Zamudio R, Ha HC. Environmental epigenetics in metal exposure. Epigenetics. 2011;6(7):820-7.
- 140. Patrick L. Toxic metals and antioxidants: Part II. The role of antioxidants in arsenic and cadmium toxicity. Altern Med Rev. 2003;8(2):106-28.
- 141. Kostova I, Balkansky S. Metal Complexes of Biologically Active Ligands as Potential Antioxidants. Curr Med Chem. 2013;20(36):4508-39.
- 142. Gaurav D, Preet S, Dua K. Chronic cadmium toxicity in rats: Treatment with combined administration of vitamins, amino acids, antioxidants and essential metals. J FDA. 2010;18(6):464-70.
- 143. Pham-Huy LA, He H, Pham-Huy C. Free radicals, antioxidants in disease and health. Int J Biomed Sci. 2008 Jun;4(2):89-96.
- 144. Tavakol HS, Akram R, Azam S, Nahid Z. Protective effects of green tea on antioxidative biomarkers in chemical laboratory workers. Toxicol Ind Health. 2013.